

Joint Dispersion Model with a Flexible Link

Rui Martins¹

April, 2016

Abstract

The objective is to model longitudinal and survival data jointly taking into account the dependence between the two responses in a real HIV/AIDS dataset using a shared parameter approach inside a Bayesian framework. We propose a linear mixed effects dispersion model to adjust the CD4 longitudinal biomarker data with a between-individual heterogeneity in the mean and variance. In doing so we are relaxing the usual assumption of a common variance for the longitudinal residuals. A hazard regression model is considered in addition to model the time since HIV/AIDS diagnostic until failure, being the coefficients, accounting for the linking between the longitudinal and survival processes, time-varying. This flexibility is specified using Penalized Splines and allows the relationship to vary in time. Because heteroscedasticity may be related with the survival, the standard deviation is considered as a covariate in the hazard model, thus enabling to study the effect of the CD4 counts' stability on the survival. The proposed framework outperforms the most used joint models, highlighting the importance in correctly taking account the individual heterogeneity for the measurement errors variance and the evolution of the disease over time in bringing new insights to better understand this biomarker-survival relation.

Keywords: Joint model, Bayesian analysis, Repeated measurements, Variance model, Time-to-event, Penalized Splines, Time-dependent coefficients.

¹email:ruimartins@egasmoniz.edu.pt

Centro de Investigação Interdisciplinar Egas Moniz (CiiEM), Escola Superior de Saúde Egas Moniz, Quinta da Granja, Monte de Caparica, 2829-511 Caparica, Portugal.

1 Introduction

A joint model is, in simple terms, an approach to the construction and analysis of the joint distribution of a set of response variables, which may be of different types. In this work we are interested in simultaneously analyse longitudinal and survival data, taking advantage of the information that flows between these two data sources collected in the same patients. The incorporation of survival information into the longitudinal process it is somewhat equivalent to take into account the effect of an informative missing-data mechanism in order to assess the trends of the repeated measures. On the other hand, include adjusted longitudinal characteristics into the survival model improves the fit of the survival regression. It is this latter feature of joint models that will permit this work to reach its objectives. We intend to perform a survival analysis of the time-to-event since the diagnostic of HIV/AIDS infected individuals using simultaneously the longitudinal information coming from the CD4⁺T lymphocyte counts (CD4 counts for short) among other covariates (*see* section 4 for a description of the dataset).

In the context of a joint analysis the dynamics of the longitudinal repeated measures are usually postulated to belong in the class of the linear mixed-effects models with Gaussian errors (Faucett and Thomas, 1996; Wulfsohn and Tsiatis, 1997; Rizopoulos, 2012). Generally it is assumed that the residual variance for the individual longitudinal trajectory is common to all subjects. Few papers on joint modelling have been published considering a model for this source of variation, which means assuming different individual residual variances (dispersion modelling). Gao et al. (2011) is an example, where the within-subject variability is assumed to have a log-normal prior distribution. McLain et al. (2012) presents a dispersion model in a frequentist framework but we can see it as an hierarchical model, easily implemented in Bayesian terms, being the variation patterns modelled as a particular function of other variables (*vide* section 2). In our study, we resorted to this strategy with promising results (section 4.4). Others authors, namely Jiang et al. (2015) assumed a latent class model for that variability and Chen and Huang (2015) and Huang et al. (2014) modelled the error process with a skew- t distribution.

One of the investigators' biggest concerns in this field is how and what characteristics should be shared by the longitudinal and survival processes. Rizopoulos and Ghosh (2011) discuss this problem by means of general families of parameterisations describing the main features of the relationship between the two processes. The most used approach is to share a set of individual random effects believed to be the basis of a latent relationship. Generally those effects are used to explain the differences between the population mean

longitudinal response and the individual-specific ones in a mixed model representation. We will extend this vision proposing the individual-specific longitudinal standard deviation as a covariate for the hazard model.

The most popular choices for the baseline hazard function specification are the parametric forms (*e.g.* Weibull - [Guo and Carlin \(2004\)](#)) or the piecewise constant approximations ([Tang et al., 2014](#); [Baghfalaki and Ganjali, 2015](#)). In seeking the most flexibility as possible we will use an approach rooted on penalized cubic B-spline functions (P-Splines for short).

The majority of the joint analysis assume that the relationship between the two processes has the same strength over time, i.e. the coefficients of the survival model accounting for the effect of a particular longitudinal characteristic are time-invariant. But if we believe, for example, that an initial significant relationship will become non-significant some time later (or *vice-versa*), what we are assuming is that the effect of the shared longitudinal characteristics on survival varies with time. We address this aspect using time-dependent coefficients approximated by P-Splines. Despite having a moderately use in survival analysis ([Hennerfeind et al. \(2006\)](#); [Hofner et al. \(2011\)](#)) time-varying coefficients are not a first option in joint models. Fortunately there are some exceptions. For instance [Yu et al. \(2014\)](#) use Splines to model these coefficients in a frequentist context; [Hanson et al. \(2011\)](#) rely on a linear combination of Gaussian kernels; [Song and Wang \(2008\)](#) use first order Taylor expansions of the function representing the time-varying coefficients.

The rest of the paper is organized as follows: section 2 describes the theoretical framework to fit a linear mixed effects model incorporating patient-specific variance, as well the survival model with time-dependent coefficients for the time to death and the linking structure; section 3 describes the estimation of this time dependent coefficients and the baseline hazard; the next (section 4) highlights the data analysis and the application of the proposed model, being the results presented and assessed. Finally we give a discussion of the paper (section 5) and supplementary material with the BUGS code to implement the models (section 6).

2 Model specification

Let us consider a study where an individual should come to an health center (hospital, clinic, laboratory, etc.) periodically in order to perform some blood tests (longitudinal process) and we follow him until he experiences the event of interest (failure) or be censored

(lost to follow-up or end of the study). The longitudinal process for the i th individual, $m_i(t)$, $i = 1, \dots, N$, is measured with error. So, the observed one is $y_i(t) = m_i(t) + e_i(t)$, $t > 0$.

The i th vector of the n_i observed repeated measures is defined as $\mathbf{y}_i = (y_i(t_{i1}), \dots, y_i(t_{in_i})) \equiv (y_{i1}, \dots, y_{in_i})$ being $\mathbf{t}_i = (t_{i1}, \dots, t_{in_i})$ a vector of fixed individual times. This encompasses the possibility of having N different measurement schedules and follow-up times. We take T_i to be the observed (possibly right censored) time-to-event, for the i th individual and δ_i to be the failure indicator. The observed data without covariates for the N independent subjects is $\mathcal{D} = \{\mathcal{D}_i\}_{i=1}^N = \{(\mathbf{y}_i, \mathbf{t}_i, T_i, \delta_i)\}_{i=1}^N$.

2.1 Longitudinal mixed dispersion model

Consider that the longitudinal outcomes, y_{ij} 's, are described by a mixed effects dispersion model (McLain et al., 2012),

$$y_{ij} | \mathbf{b}_i, \sigma_i^2 \sim \mathcal{N}(m_i(t_{ij}), \sigma_i^2), \quad j = 1, \dots, n_i \quad (1)$$

$$m_i(t_{ij}) = \boldsymbol{\beta}_1^\top \mathbf{x}_{1i}(t_{ij}) + \mathbf{b}_{1i}^\top \mathbf{w}_{1i}(t_{ij}), \quad (2)$$

$$\sigma_i^2 = \sigma_0^2 \exp\{ \boldsymbol{\beta}_2^\top \mathbf{x}_{2i}(t_{ij}) + \mathbf{b}_{2i}^\top \mathbf{w}_{2i}(t_{ij}) \}, \quad (3)$$

where \mathbf{x}_{1i} , \mathbf{x}_{2i} , \mathbf{w}_{1i} and \mathbf{w}_{2i} are appropriate subject-specific vectors of covariates (possibly time-dependent); $\boldsymbol{\beta}_1$ and $\boldsymbol{\beta}_2$ are vectors of population regression parameters; $(\mathbf{b}_{1i}^\top, \mathbf{b}_{2i}^\top) = \mathbf{b}_i | \Sigma \sim \mathcal{N}_p(\mathbf{0}, \Sigma)$ are time-independent subject-specific random effects capturing the inherent biological variability between individuals and allowing for between-individual heterogeneity in the mean and variance.

One of the common and major assumptions in longitudinal models is that the within-individual variances are homogeneous, although this is not always satisfied. For example, considering our dataset (section 4.1), the plot of the individual $\sqrt{\text{CD4}}$ values against the standard deviation (Figure 1) suggests considerable within-subject variance heterogeneity. Large values of the mean are associated with high variability. In equation (3) the residual variance, σ_i^2 , is assumed to be an individual property allowing for heterogeneity in the variance trends among the individuals. Modelling it and identify covariates related to this discrepancies seems wise. Particularly, in many HIV/AIDS studies, where investigators are interested in understanding the trends of the variability, having an individual estimate of the subject-residual variance can be a plus in the assessment of whether individuals with different biomarker's stability have different prognosis.

The representation in (3), where σ_0^2 acts as a “baseline” variance, allows for both increasing and decreasing variance across individuals. \mathbf{x}_{2i} and \mathbf{w}_{2i} represent the variables influencing the within-subjects variance. In this way, one can examine whether contextual variables are related to the within-individual variance. In addition, this specification can be considered as an extension to the dispersion model defined in Lin et al. (1997) or the one defined by Gao et al. (2011). The former modelled the individual-specific measure of stability, σ_i^2 , through an inverse gamma prior distribution, which is a special case of (3). The latter do not consider possible covariate effects. Indeed, if we consider $\exp\{\boldsymbol{\beta}_2^\top \mathbf{x}_{2i}(t_{ij}) + \mathbf{b}_{2i}^\top \mathbf{w}_{2i}(t_{ij})\} = 1$ in (3) we have $\sigma_i^2 = \sigma_0^2$ and we are back to the simple models where σ_i^2 accounts for the randomness in stability by using some prior distribution for this parameter. More details about the choose of this prior distribution are discussed in section 4.3.

2.2 Hazard model with time-varying coefficients

Various approaches have been proposed to link the longitudinal and survival processes. In this work, we tackle joint modelling considering the inclusion of some characteristics of the longitudinal trajectory, namely the random effects, \mathbf{b}_i , and the standard deviation, σ_i , into a hazard model (shared parameters approach), described below,

$$h_i(t \mid \mathbf{b}_i, \sigma_i) = h_0(t) \exp\{\boldsymbol{\beta}_3^\top \mathbf{x}_{3i} + \mathcal{C}_i\{\mathbf{b}_i, \sigma_i; \mathbf{g}(t)\}\} = h_0(t) \exp\{\varrho_i(t)\}, \quad (4)$$

where \mathbf{x}_{3i} is a subject-specific vector of baseline covariates and $\boldsymbol{\beta}_3$ is the respective population parameters vector. $\mathcal{C}_i\{\cdot\}$ is a function specifying which components of the longitudinal process are directly related to $h_i(\cdot)$. Finally, $\mathbf{g}(t) = (g_1(t), \dots, g_L(t))$ is an appropriate vector, set case-by-case, of L smooth functions (approximated by P-Splines - see section 3.1) representing the time-varying regression coefficients (Hennerfeind et al., 2006), which measure the effect of some particular characteristics of the longitudinal outcome to the hazard. These coefficients are particularly useful in explaining the effect of a time-independent covariate on survival when its impact is not constant throughout the time, *e.g.* if an initially significant relation becomes non-significant after a certain period. Baseline hazard, $h_0(t)$, can have a parametric form (*e.g.* Weibull) or be specified using P-Splines or a piecewise constant function.

3 Estimation, specification and model choice

In this section we present the estimation of the time-varying coefficients, $g_l(t)$, $l = 1, \dots, L$, the different forms that the baseline hazard function, $h_0(\cdot)$, can have and the specification of $\mathcal{C}_i\{\cdot\}$, as well as the posterior distribution of the joint model. Finally, we will allude to the Watanabe-Akaike information criterion (WAIC, [Watanabe \(2010\)](#)) for model choice.

3.1 Time-varying coefficients

To fit model (4) we will apply a penalized spline regression method to the elements of the vector $\mathbf{g}(t)$. The main idea of a regression via splines is described as follows and more details can be found in [Lang and Brezger \(2004\)](#), [Brezger and Lang \(2006\)](#) and [Hennerfeind et al. \(2006\)](#). Using a linear combination of Q_l B-spline basis functions, $\{B_{lq}(t)\}_{q=1}^{Q_l}$, $l = 1, \dots, L$, defined over a grid of $s_l + 1$ equally spaced knots, $t_{\min} = \kappa_{l0} < \kappa_{l1} < \dots < \kappa_{ls_l} = t_{\max}$, over the domain of t , we will be able to well approximate $g_l(t)$. As in [Eilers and Marx \(2010\)](#) all the basis functions have the same shape including the boundaries of the domain, contrasting, for example, with natural B-splines, where near the boundaries the basis functions have different shapes. In our case the splines are of order $p = 3$ and computed as differences of truncated power functions ([Eilers and Marx, 2010](#)). Thus, the expansion of $g_l(t)$ into B-spline basis is

$$g_l(t) = \sum_{q=1}^{Q_l} \gamma_{lq} B_{lq}(t), \quad l = 1, \dots, L, \quad (5)$$

where $Q_l = s_l + 3$, $\boldsymbol{\gamma}_l = (\gamma_{l1}, \dots, \gamma_{lQ_l})$ are unknown vectors of fixed coefficients and $B_{lq}(t)$ denotes the q th basis function of the B-Spline. From now on we will drop index l from the notation of the Q_l , κ_l , s_l , and $B_{lq}(t)$, because we are assuming the same basis for all the B-Splines associated with each $g_l(t)$. To ensure that our functions $g_l(t)$ are smoothed enough, we penalize the first order differences between adjacent elements in $\boldsymbol{\gamma}_l$ using a first order random-walk to define their priors, i.e.,

$$\gamma_{l1} \sim \mathcal{N}(0, 1000), \quad \gamma_{lq} \mid \tau_l^2 \sim \mathcal{N}(\gamma_{l,q-1}, \tau_l^2), \quad l = 1, \dots, L; \quad q = 2, \dots, Q, \quad (6)$$

where τ_l^2 acts as the variance controller. Large values correspond to a rougher curve. First order penalty aims to avoid abrupt jumps between adjacent elements in $\boldsymbol{\gamma}_l$. Instead a second order random-walk could be used to avoid departures from a linear trend as in [Lang and Brezger \(2004\)](#) and [Kneib and Fahrmeir \(2007\)](#).

Joint prior of the elements in γ_l is defined as a product of conditional densities defined by (6) yielding a Multivariate Gaussian distribution

$$\pi(\gamma_l | \tau_l^2) \propto \tau_l^{-(Q-1)} \exp \left\{ -\frac{1}{2\tau_l^2} \gamma_l^\top \mathbf{P} \gamma_l \right\}, \quad l = 1, \dots, L; \quad q = 1, \dots, Q. \quad (7)$$

$\mathbf{P} = \mathbf{D}^\top \mathbf{D}$ is the precision matrix and \mathbf{D} is the $(Q-1) \times Q$ matrix of the first order differences.

3.2 Baseline hazard

Various choices have been made to deal with $h_0(t)$. In the traditional Cox model (Cox, 1972) it is treated as a nuisance parameter and left unspecified, relying the estimation process on the partial-likelihood function (Costa and Shaw, 2009). However, a completely unspecification can cause underestimation (Hsieh et al., 2006). On the other hand we have those who completely specify it using a parametric form (*e.g.* Weibull) as in Guo and Carlin (2004). Other popular choices are specifications by splines (Rizopoulos et al. (2009) and Kneib and Fahrmeir (2007)) or a piecewise constant function (Tseng et al. (2005), Zhu et al. (2012) and Rizopoulos and Ghosh (2011) just to name a few). A baseline specification provides the possibility to direct estimate the full survival time distribution contrary to what happens when we left $h_0(t)$ unspecified.

Assuming that each T_i , $i = 1, \dots, N$, has a Weibull distribution, $\mathcal{W}(\rho, e^{\varrho_i(t)})$, $h_i(t|\cdot)$ in (4) is defined as

$$h_i(t | \mathbf{b}_i, \sigma_i) = \rho t^{\rho-1} \exp\{\varrho_i(t)\}. \quad (8)$$

In case of a spline approach $h_i(t|\cdot)$ is defined as

$$h_i(t | \mathbf{b}_i, \sigma_i) = \exp\{g_0(t) + \varrho_i(t)\}, \quad (9)$$

where now $\mathbf{g}(t) = (g_0(t), g_1(t), \dots, g_L(t))$ accommodates $g_0(t) = \log[h_0(t)]$, which is approximated in a similar manner to that of the $g_l(t)$ coefficients, $l = 1, \dots, L$, in section 3.1. Finally, if $h_0(t)$ is a piecewise constant function defined over a finite partition of the time, $0 = a_1, a_2, \dots, a_K, a_{K+1} = \infty$, we define the baseline hazard as $h_0(t) = \sum_{k=1}^K \lambda_k \mathbb{1}_{[a_k, a_{k+1}[}(t)$, where $\boldsymbol{\lambda} = (\lambda_1, \dots, \lambda_K)$ is a vector of positive but unknown parameters, each one representing the constant local hazard for $t \in [a_k, a_{k+1}[$, $k = 1, \dots, K$. Thus we can write

$$h_i(t | \mathbf{b}_i, \sigma_i) = \sum_{k=1}^K \lambda_k \mathbb{1}_{[a_k, a_{k+1}[}(t) \exp\{\varrho_i(t)\}. \quad (10)$$

3.3 The posterior distribution

To form the contribution of the i th individual to the likelihood we assume a non-informative right censoring, events and censoring are independent of the process measurement schedule, the longitudinal and survival processes are independent given the random effects, \mathbf{b}_i , and, in addition, the elements of the vector \mathbf{y}_i are assumed independent given the same set of random effects. Thus, the posterior distribution, $\pi(\boldsymbol{\theta}|\mathcal{D})$, will be proportional to

$$\begin{aligned} & L_{1i}(\boldsymbol{\theta} \mid \mathcal{D}_i) \times L_{2i}(\boldsymbol{\theta} \mid \mathcal{D}_i) \times \pi(\boldsymbol{\theta}) \\ &= \left\{ \prod_{j=1}^{n_i} f(y_{ij}|\mathbf{b}_i, \sigma_i^2) \right\} \times \left\{ h(T_i|\mathbf{b}_i)^{\delta_i} S(T_i|\mathbf{b}_i) \right\} \times \pi(\boldsymbol{\theta}), \end{aligned} \quad (11)$$

where $\pi(\boldsymbol{\theta})$ denotes the prior distribution of the full parameters vector including the random effects, \mathbf{b}_i ; $L_{1i}(\boldsymbol{\theta}|\mathcal{D}_i)$ is the i th individual longitudinal contribution, being $f(\cdot)$ the Gaussian probability density function specified as

$$f(y_{ij}|\mathbf{b}_i, \sigma_i^2) = \frac{1}{\sqrt{2\pi\sigma_i^2}} \exp \left\{ -\frac{[y_{ij} - m_i(t_{ij})]^2}{2\sigma_i^2} \right\}, \quad (12)$$

and finally $h(\cdot)$ and $S(\cdot)$ are, respectively, the hazard and the survival function for T_i .

All of our joint models will have the same longitudinal specification (1). On the other hand, hazard models (8), (9) and (10) will define three different joint models, implying three different specifications for the i th individual survival contribution, $L_{2i}(\boldsymbol{\theta}|\mathcal{D}_i)$. Thus, considering (8)

$$L_{2i}(\boldsymbol{\theta} \mid \mathcal{D}_i) = \{\rho T_i^{\rho-1} e^{\varrho_i(T_i)}\}^{\delta_i} \exp\{-e^{\varrho_i(T_i)} T_i^\rho\}, \quad (13)$$

assuming (9)

$$L_{2i}(\boldsymbol{\theta} \mid \mathcal{D}_i) = \{e^{g_0(T_i) + \varrho_i(T_i)}\}^{\delta_i} \exp\left\{-\int_0^{T_i} e^{g_0(u) + \varrho_i(u)} du\right\}, \quad (14)$$

and finally, postulating (10) we have

$$L_{2i}(\boldsymbol{\theta} \mid \mathcal{D}_i) = \{\lambda_{\mathring{k}} e^{\varrho_i(T_i)}\}^{\delta_i} \exp\left\{-\left[\lambda_{\mathring{k}}(T_i - a_{\mathring{k}}) + \sum_{k=1}^{\mathring{k}-1} \lambda_k(a_{k+1} - a_k)\right] e^{\varrho_i(T_i)}\right\}, \quad (15)$$

where \mathring{k} is the largest integer such that $a_{\mathring{k}} \leq T_i$.

Although we are working on a Bayesian framework we still have to evaluate the integral in (14). In our models we used a 15-points Gauss-Kronrod quadrature rule as in Rizopoulos and Ghosh (2011), but other numeric methods could be applied. Ibrahim et al. (2004) show an alternative to this approximation tool.

3.4 Model choice

The purposes of model comparison and selection will be achieved with a recent measure. The Watanabe-Akaike information criterion (WAIC, [Watanabe \(2010\)](#)), or the widely applicable information criterion in its own words, is a measure of predictive accuracy and invariant to reparameterisations contrary to the DIC ([Spiegelhalter et al., 2002](#)). In addition WAIC is fully Bayesian and closely approximates Bayesian cross-validation. DIC is considered not fully Bayesian, in part because it is based on a point estimate. Following [Gelman et al. \(2014\)](#) we define the WAIC value as

$$\begin{aligned} \text{WAIC} &= -2 [\text{lppd} - p_{\text{WAIC}}] \\ &= -2 \left[\log \left\{ \prod_{i=1}^N p(\mathcal{D}_i | \mathcal{D}) \right\} - \sum_{i=1}^N \mathbb{V}_{\boldsymbol{\theta} | \mathcal{D}} [\log \{p(\mathcal{D}_i | \boldsymbol{\theta})\}] \right]. \end{aligned} \quad (16)$$

Like the DIC models with smaller WAIC values should be preferred. More details about this criterion, including the computation in the MCMC context, can be found in the SuppMaterial.

4 HIV/AIDS data analysis

4.1 Dataset

Brazilian National AIDS Program generated three major electronic databases ([Fonseca et al., 2010](#)): (i) SINAN-AIDS (Information System for Notifiable Diseases of AIDS Cases) which is the most important electronic AIDS surveillance database, with all cases reported since 1980; (ii) SISCEL (Laboratory Test Control System) designed to monitor laboratory tests, such as CD4 counts and viral load tests for HIV/AIDS patients followed in the public health sector since 2002; (iii) SICLOM (System for Logistic Control of Drugs) developed to control the logistic for the AIDS treatment deliveries; it shares the patients list with SISCEL since 2002. These three databases have been previously combined in a single database with both HIV and AIDS cases using a process called record linkage, which was adopted by the Surveillance Unit of the Brazilian National AIDS Program ([Fonseca et al., 2010](#)). This linkage strategy has been increasingly used in AIDS surveillance and research ([Deapen et al., 2007](#)) to verify under-reporting of cases and eliminate the duplicated ones. In Brazil, that procedure has improved the quality of HIV/AIDS data information ([Fonseca et al., 2010](#)). Notice that 2002–2006 can be considered as the

first period with substantial information on both HIV/AIDS survival and CD4 exams, where 88 laboratories located in all 27 Brazilian states were using SISCEL, covering 90% of all CD4 and viral load exams carried out by the public health sector. Cases diagnosed before 2002 were excluded because personal identifiers were not available in the mortality database for the entire country before that date (Fonseca et al., 2010). For institutional reasons, we had access only to a simple random sample of the combined database.

The longitudinal and survival data were collected in a network of 88 laboratories located in every 27 states of Brazil during the years 2002–2006. CD4 counts (a measure of immunologic and virologic status) and survival time after diagnosis were the responses collected in a random sample of $N = 500$ individuals, under HAART therapy, from the original data base. While the explanatory variables included were: **age** ($[15, 50[$, coded 0; ≥ 50 , coded 1), **gender** (Female, coded 0; Male, coded 1), **PrevOI** (previous opportunistic infection at study entry, coded 1; no previous infection, coded 0), date of HIV/AIDS diagnosis and date of death (available if death happened before 31st December 2006 and censored otherwise). The survival time after diagnosis is calculated as the time period, in years, between date of diagnosis and date of death. As referred in Souza-Jr et al. (2007) the variable that accounts for age was chosen basis on the Ministry of Health recommendations, as the over-50 age group showed a higher proportion of delayed initiation of therapy when compared to the population group aged 15-49 years.

There were 34 deaths in this dataset; 440 (88%) of the patients were between 15 and 49 years old; 298 (59.6%) were males of whom 23 died. 302 (60.4%) individuals had no previous infection; 6.4% lived in the Central-West, 9.6% in the North-east, 4.6% in the North, 64.8% in the South-east region and 14.6% in the South. The initial median CD4 count was 269 cells/mm³ (men - 250 cells/mm³; women - 295 cells/mm³) and patients made on average 5.51 CD4 exams resulting in a total of 2757 observations. The mean time of a censored patient in the study was 930.2 days (approximately 2.56 years) and for those which had the event that mean time was 862.4 days (approximately 2.36 years).

4.2 Fitted models

Our main concern in this work is to use joint modelling for making appropriate inferences about the influence of the longitudinal data characteristics on the survival time. Table 1 shows the form of the 33 joint models fitted to the data and the respective WAIC values for comparison. A square root transformation of the CD4, $\sqrt{\text{CD4}}$, was used in the analysis in order to deal with the high degree of skewness towards the higher counts. Its adjusted

longitudinal mean response, $m_i(t_{ij})$, has always the following representation

$$m_i(t_{ij}) = \beta_{10} + \beta_{11}t_{ij} + \beta_{12}\mathbf{sex}_i + \beta_{13}\mathbf{age}_i + \beta_{14}\mathbf{Prev0I}_i + b_{1i,1} + b_{1i,2}t_{ij}, \quad (17)$$

where $\boldsymbol{\beta}_1^\top = (\beta_{10}, \dots, \beta_{14})$, $\mathbf{b}_{1i}^\top = (b_{1i,1}, b_{1i,2})$, $\mathbf{x}_{1i}(t_{ij}) = (1, t_{ij}, \mathbf{sex}_i, \mathbf{age}_i, \mathbf{Prev0I}_i)$ and $\mathbf{w}_{1i}(t_{ij}) = (1, t_{ij})$.

The dispersion model in (3) will assume several formulations. Supposing that a set of baseline covariates are related to the variability and defining $\boldsymbol{\beta}_2^\top = (\beta_{21}, \beta_{22}, \beta_{23})$, $\mathbf{b}_{2i}^\top = b_{2i}$, $\mathbf{x}_{2i}(t_{ij}) = (\mathbf{sex}_i, \mathbf{age}_i, \mathbf{Prev0I}_i)$ and $\mathbf{w}_{2i}(t_{ij}) = 1$, we can use the form below to study its effects in the CD4 stability,

$$\sigma_i^2 = \sigma_0^2 \exp\{\beta_{21}\mathbf{sex}_i + \beta_{22}\mathbf{age}_i + \beta_{23}\mathbf{Prev0I}_i + b_{2i}\}, \quad (18)$$

where σ_0^2 is the population's common variance and b_{2i} captures different kinds of heterogeneity between individuals, in the same spirit as the frailties in a hazard model. If ones assume that these random effects, b_{2i} , are enough to explain the heteroscedasticity, we can define

$$\sigma_i^2 = \sigma_0^2 \exp\{b_{2i}\}. \quad (19)$$

More simple forms can be tested, namely considering an individual error variance, σ_i^2 , to which we assign an exchangeable prior distribution, i.e.

$$\sigma_i^2 | \boldsymbol{\theta}_{\sigma^2} \sim \pi(\boldsymbol{\theta}_{\sigma^2}), \quad (20)$$

which means that individual variance, σ_i^2 's, are i.i.d. random variables conditional on the unknown hyperparameters $\boldsymbol{\theta}_{\sigma^2}$. A common error variance can also be assumed, with prior distribution

$$\sigma_i^2 = \sigma_0^2 \sim \pi(\boldsymbol{\theta}_{\sigma^2}), \quad i = 1 \dots, N. \quad (21)$$

In the survival part we will always consider the hazard model defined in (4) taking

$$\boldsymbol{\beta}_3^\top \mathbf{x}_{3i} = \beta_{31}\mathbf{sex}_i + \beta_{32}\mathbf{age}_i + \beta_{33}\mathbf{Prev0I}_i, \quad (22)$$

conjugated with three forms for the function $\mathcal{C}_i(\cdot)$. First a linear one with three individual-specific random effects

$$\mathcal{C}_i(\cdot) = g_1(t)b_{1i,1} + g_2(t)b_{1i,2} + g_3(t)b_{2i}, \quad (23)$$

which is the traditional shared random effects approach ([Henderson et al. \(2000\)](#)) extended with b_{2i} , which carries information about the individual-specific variability. A form accounting for the explicit effect of the individual longitudinal standard deviation

$$\mathcal{C}_i(\cdot) = g_1(t)b_{1i,1} + g_2(t)b_{1i,2} + g_3(t)\sigma_i, \quad (24)$$

which is an approach inspired in [Gao et al. \(2011\)](#) and [McLain et al. \(2012\)](#). Finally a structure without any information about the residual variability

$$\mathcal{C}_i(.) = g_1(t)b_{1i,1} + g_2(t)b_{1i,2}. \quad (25)$$

Equations (23) and (24) states that there is some information about the survival process coming from σ_i^2 , i.e. the residual variance may be an indicator of an increase or decrease in the hazard of death. For the time-varying coefficients, $g_1(t)$, $g_2(t)$ and $g_3(t)$ we will use 21 knots (i.e. $s = 20$ and $Q = 23$) which corresponds to a knot every 3 months during the 5 years. The same is true for $g_0(t)$ when used in (9). When a piecewise constant baseline-hazard is used, and in order to have a term of comparability with $g_0(t)$, we used 20 subintervals, $K = 21$, with equal lengths corresponding also to a length of 3 months each during the 5 years.

4.3 Prior specification

Under the Bayesian framework we need to inform about the prior specifications for all the unknown parameters. We assume that all the elements of the vectors β_1 , β_2 and β_3 are independent of each other and Gaussian distributed, $\mathcal{N}(0, 100)$. A Wishart prior distribution, $Wish(R, \xi)$, was set as the prior for the inverse of the variance-covariance matrix, Σ^{-1} , of the random effects, \mathbf{b}_i , where $R = \text{diag}(100)$ and $\xi = 500/20$ to avoid confounding between the fixed and random effects ([Carlin and Louis \(2001\)](#), p.277). Priors for γ_l , $l = 0, 1, 2, 3$, are defined in (6). Generally we will assign a Gamma prior to the inverse of the smoothing parameter, $1/\tau_l^2 \sim \mathcal{G}(0.001, 0.001)$, $l = 0, 1, 2, 3$.

When the hazard model is defined according to (10) we should use some prior specification for the elements of λ , which are unknown. The prior here considered will allow to center the baseline hazard on an Exponential distribution, which has constant hazard, by defining independent Gamma distributions, $\mathcal{G}(\epsilon_{\lambda_k}, \epsilon_{\lambda_k})$, on the λ_k 's, implying that $\mathbb{E}[\lambda_k] = 1$, i.e. our initial guess is that each piece of the baseline hazard function is constant and equal to 1. We will fix $\epsilon_{\lambda_k} = 0.001$, but an hyperprior distribution could be placed on it. Although we should be careful because we can easily end with an over-fitted model. Thus we assume $\lambda_k \sim \mathcal{G}(0.001, 0.001)$, $k = 1, \dots, K$. Using small values for ϵ_{λ_k} means larger prior variance for λ_k allowing the data to drive departures from the Exponential hazard.

Concerning the dispersion model in (18) if the linear predictor $\eta_i = \{\beta_{21}\text{sex}_i + \beta_{22}\text{age}_i + \beta_{23}\text{PrevOI}_i + b_{2i}\} = 0$, then $\sigma_i^2 = \sigma_0^2$ and all the individuals have the same residual variance. Instead if $\eta_i \neq 0$ then we are assuming the existence of some differences in the residual

variance behaviour across individuals. Our choice to define the prior for the variance parameter, σ_0^2 , in the scenarios (18), (19) and (21) is an approximation to the Jeffreys prior, $\log(\sigma_0) \sim \mathcal{U}(-A, A)$ with A large (100 in this case). The same prior distribution is considered for the individual error variance, σ_i^2 , in (20). Another two different priors were considered, namely the traditional, $1/\sigma_0^2 \sim \mathcal{G}(\epsilon, \epsilon)$ and a half-Cauchy distribution, $\sigma_0|\varpi \sim \text{h-}\mathcal{C}(\varpi)$, $\varpi \sim \mathcal{U}(0, 100)$. This latter approximation is a natural option when some variances may be outlying (Gelman, 2006). It also facilitates the specification of prior beliefs because the only parameter represents the population median standard deviation (Lunn et al. (2013), p.237). To sample from a half-Cauchy distribution in WinBUGS we have to resort in some distributional calculations, because it's not available in a standard form (for more details *vide* Lunn et al. (2013), p.225). Although in the results section we only present findings considering that $\log(\sigma_0) \sim \mathcal{U}(-A, A)$, because the conclusions based on the WAIC values and the posterior estimates for the fitted models (Tables 1 and 2) showed no evidence of being affected by that prior choice.

4.4 Results

Assuming that all the elements of $\boldsymbol{\theta}$ are independent of each other, model fitting was performed through MCMC simulation within WinBUGS (Lunn et al., 2000) by specifying the full likelihood function and the prior distributions for the unknown parameters. The obtained estimates were based on chains of 500,000 iterations following 250,000 iterations of “burn-in”. In order to eliminate autocorrelation among samples within the chains, we selected every 25th iteration of the chains. A study of the trace and density plots of the posterior distributions indicated no convergence problems concerning these samples.

To generate the likelihood when using a piecewise constant function to approximate the baseline hazard as in (15) we use a “trick” that relies on Poisson modelling of expanded data, which means that we are going from an infinite dimensional and unspecified, $h_0(t)$, to a K dimensional parameter vector, $\boldsymbol{\lambda} = (\lambda_1, \dots, \lambda_K)$. We will not discuss the details here because going into these issues would distract from our main purpose. There exists a broad list of references that can be checked to deepen this subject. The reader should for example consult Christensen et al. (2011, p. 347) for the theoretical questions and for the WinBUGS implementation Lunn et al. (2013, p. 289). Details about the prior specifications for all the parameters in $\boldsymbol{\theta}$ will be given in section 4.3.

Based on the smallest WAIC value (Table 1), a P-Spline approach to the baseline hazard seems to be the best strategy. The chosen model \blacktriangle ($\sigma_i^2 = (20)$; $\psi_i = (22) + (24)$) assumes a

heterogeneous within subject variability considering an exchangeable prior distribution for the σ_i 's. Additionally, the patient-specific random intercept, slope and standard deviation are considered as covariates for the hazard model. The inclusion of the subject-specific variability as a covariate to explain the survival outcome ($\psi_i = (22) + (24)$) improves the results (lower WAIC) compared to the models without this feature ($\psi_i = (22) + (23)$ or $\psi_i = (22) + (25)$). This is scientifically appealing, because the CD4's stability might contain extra information about health-related risks, complementing that provided by the mean-level trajectory. Although, for this particular case, our initial feeling – a possible effect of some set of covariates related to the individual heterogeneity – did not show up (models \diamond ; $\sigma_i^2 = (18)$).

The “traditional model” to jointly analyse longitudinal and survival data (Faucett and Thomas, 1996; Henderson et al., 2000) considers: (i) a common residual variance as in (21), (ii) a set of two shared random effects ($b_{1i,1}$ and $b_{1i,2}$) believed to drive the relation between the two processes and (iii) the coefficients accounting for the effects of these latent variables on the hazard are time-independent, i.e., g_1 and g_2 do not depend on t , being the function that specifies which components of the longitudinal process are shared with the survival one defined as $\mathcal{C}_i(.) = g_1 b_{1i,1} + g_2 b_{1i,2}$. In the most part of the literature, the baseline hazard has been tackled considering a Weibull form, but approaches considering splines or a piecewise-constant approximation also have its place (Rizopoulos and Ghosh, 2011). Last row of Table 1 has been added so we can compare the performance of these “traditional” approaches (models \diamond) with our proposed models, namely \triangle , \square and \circ . Considering only the situations where h_0 is adjusted using P-Splines, we note that the traditional model \blacklozenge is the worst in terms of its WAIC value.

Table 2 shows the posterior mean estimates together with the 95% Credible Interval (CI) for a set of parameters for the best model \blacktriangle compared to its counterparts for the traditional model \blacklozenge . One can see that the variables **gender**, **age** and **PrevO1** are all significant in explaining the different longitudinal trajectories and survival times. Older males with opportunistic infections at the study entry have lower $\sqrt{\text{CD4}}$ values (negative estimates). Younger females without opportunistic infections at the study entry have a lower hazard of death (positive estimates). Model \blacktriangle has been able to capture the effect of the **gender** on the survival outcome as opposed to the model \blacklozenge . There is another important thing coming out of these estimates. The unique elements, σ_{11}^b , σ_{12}^b and σ_{22}^b , of the random-effects covariance matrix, Σ , for the chosen model are lowered compared to its counterparts for the model \blacklozenge , which means that we have been able to shrink more the individual random-effects under the presence of a dispersion model for

the individual-specific standard deviation than considering a common within-individual variance. This happens because the dispersion model is now explaining some of that additional variability. Some authors, namely [Lyles et al. \(1999\)](#), have already noted this feature – random within-subject variances appears to describe a collection of longitudinal biomarker measurements well, as compared to a broad range of alternative models for the correlation structure.

Figure 2 displays the histogram for the posterior estimates of the within-individual standard deviation, σ_i , which are considerably heterogeneous and right skewed. This fact is one more reason supporting the assumption of heterogeneous variance which led us to consider a dispersion model when analysing the CD4 repeated measures. These estimates contrast with the common value, $\sigma_0 = 2.59$, for the model \blacklozenge where a structure for the dispersion is not considered.

Figure 3 top left panel presents $g_0 = \log(h_0)$. Because its posterior mean (black line) is approximately horizontal and the 95% Credible Band (CB) lies, approximately, in $(-1, -3)$, we can consider that HIV/AIDS individuals taking medicines correctly have a lower baseline hazard and do not suffer major changes over time. This plot seems to tell us that nowadays people living with HIV, if correctly treated, will have a large life expectancy. Indeed, [May et al. \(2014\)](#) have already noted this in a big study involving 21,388 patients in the UK. Some groups of HIV-positive individuals may expect to live a similar life span to that of the general population, others have reduced life expectancy due to the impact of late diagnosis and late initiation of the therapy. Unfortunately our dataset does not have information about the elapsed time since the infection until the treatment beginning. We only know that older people take longer to start the therapy. The top right panel shows that initially the individual random intercept, $b_{1i,1}$, does not have a significant effect on the hazard but after a short period of time, less than a year, individuals with CD4 counts at the study entry above the mean are at lower risk until, approximately, the begin of the third year when that initial value becomes negligible again. Indeed after a certain period of time it is expected that the initial values become less important. The bottom left panel shows that an individual with a positive time trend for the biomarker has a decreased hazard of death up to 3.5 years after diagnostic, approximately, because the 95% CB lies below the horizontal dotted line, $g_2(t) = 0$. After that period the impact is lowered. Bottom right panel suggests that the higher the within-individual variability the higher the hazard of death and that effect is growing over the time, albeit only after 1.5 years approximately, because the horizontal dotted line, $g_3(t) = 0$, is in the 95% CB before that period. Biologically this is understandable, because if after 1 or 2 years

the CD4 counts variability doesn't stabilize, that means that patient's immune system is responding poorly to the treatment.

Have the possibility to incorporate time-dependent coefficients into a model is often a bonus. Without it we could not have given the interpretations to the coefficients \mathbf{g} that we gave above. Actually, the posterior estimates for g_1 and g_2 in model \blacklozenge are, respectively, -0.21 and -0.54 , which leads us to gave the following explanation: during the patients follow-up, individuals with higher CD4 values at the study entry and with an upward trend for the CD4 trajectory have a decrease in their risk of death. But, due to the fact that our chosen model \blacktriangle has time-dependent g 's we know now that after a certain period of time both the initial value and the trajectory trend become non-significant.

Unlike the random effects, \mathbf{b}_i , the biomarker variability, σ_i , has an intuitive meaning for the physicians and respective patients. In a joint model context its interpretation is straightforward, because its contribution to understand the time-to-event can be readily quantified as a hazard ratio (HR). For instance, considering two subjects only differing in their repeated measures variability, everything else being equal for a specific time, the hazard ration is $\text{HR} = \exp\{g_3(t)\}$, which is familiar to the clinical staff.

5 Discussion

This paper explores the association between the CD4's trajectory with the time-to-death for patients diagnosed with HIV/AIDS and under HAART therapy, which has important implications in the disease comprehension. The three new aspects presented in this work seem to be improving the traditional framework, videlicet: the dispersion model is capturing some extra-variability in the individual repeated measures implying a reduction in the variability of the individual-specific random effects; the use of the individual level standard deviation as a covariate in the hazard model is scientifically appealing, highlighting that the biomarker's stability might contain extra information about health-related risks, complementing that provided by the mean-level trajectory; the time-dependent coefficients, estimated via P-Splines, that account for the link between the two processes, allow us to understand the influence of the biomarker's values and its variability throughout the time in the survival-related outcome. Obviously, all these results need more studies to support them, but they seem to be encouraging. Beyond AIDS framework, there is an enormous potential to apply the methods developed in this work to other epidemiological contexts.

Acknowledgements

The author wish to thank Maria Goretti Fonseca and Cláudia Coeli for the database. This work was partially funded by Egas Moniz, Cooperativa de Ensino Superior.

References

- Baghfalaki, T. and M. Ganjali (2015, June). A bayesian approach for joint modeling of skew-normal longitudinal measurements and time to event data. *REVSTAT-Statistical Journal* 13(2), 169–191.
- Brezger, A. and S. Lang (2006). Generalized structured additive regression based on bayesian p-splines. *Computational Statistics and Data Analysis* 50, 967–991.
- Carlin, B. and T. Louis (2001). *Bayes and Empirical Bayes Methods for Data Analysis, Second Edition*. Chapman & Hall.
- Chen, J. and Y. Huang (2015, September). A Bayesian mixture of semiparametric mixed-effects joint models for skewed-longitudinal and time-to-event data. *Statistics in Medicine* 34(20), 2820–2843.
- Christensen, R., W. Johnson, A. Branscum, and T. E. Hanson (2011). *Bayesian ideas and data analysis: An introduction for scientists and statisticians*. Boca Raton, Chapman & Hall/CRC Press. ISBN:978-1-4398-0354-7.
- Costa, M. J. and J. E. H. Shaw (2009, Jan). Parametrization and penalties in spline models with an application to survival analysis. *Computational Statistics & Data Analysis* 53(3), 657–670.
- Cox, D. (1972). Regression models and life tables. *Journal of the Royal Statistical Society Series B* 34, 187–220.
- Deapen, D., M. Cockburn, R. Pinder, S. Lu, and A. Wohl (2007, Aug). Population-based linkage of AIDS and cancer registries. *American Journal of Preventive Medicine* 33(2), 134–136.
- Eilers, P. H. C. and B. D. Marx (2010). Splines, knots, and penalties. *Wiley Interdisciplinary Reviews: Computational Statistics* 2(6), 637–653.
- Faucett, C. L. and D. C. Thomas (1996, Aug). Simultaneously modelling censored survival data and repeatedly measured covariates: a gibbs sampling approach. *Statistics in Medicine* 15(15), 1663–1685.
- Fonseca, M. G., C. M. Coelli, F. Lucena, V. Veloso, and M. Carvalho (2010). Accuracy of a probabilistic record linkage strategy applied to identify deaths among cases reported to the brazilian aids surveillance database. *Cad. Saúde Pública* 26, 1431–1438.
- Gao, F., J. Miller, C. Xiong, J. Beiser, and M. Gordon (2011). A joint-modeling approach to assess the impact of biomarker variability on the risk of developing clinical outcome. *Statistical Methods & Applications* 20, 83–100.
- Gelman, A. (2006). Prior distributions for variance parameters in hierarchical models. *Bayesian Analysis* 1(3), 1–19.

- Gelman, A., J. Hwang, and A. Vehtari (2014). Understanding predictive information criteria for bayesian models. *Statistics and Computing* 24(6), 997–1016.
- Guo, X. and P. Carlin (2004). Separate and joint modelling of longitudinal and event time data using standard computer packages. *The American Statistician* 58(1), 16–24.
- Hanson, T., A. Branscum, and W. Johnson (2011). Predictive comparison of joint longitudinal-survival modeling: a case study illustrating competing approaches. *Lifetime Data Analysis* 17, 3–28. 10.1007/s10985-010-9162-0.
- Henderson, R., P. Diggle, and A. Dobson (2000, Dec). Joint modelling of longitudinal measurements and event time data. *Biostatistics* 1(4), 465–480.
- Hennerfeind, A., A. Brezger, and L. Fahrmeir (2006, 09). Geoadditive survival models. *JASA* 101, 1065–1075.
- Hofner, B., T. Kneib, W. Hartl, and H. Küchenhoff (2011, Jan). Building cox-type structured hazard regression models with time-varying effects. *Statistical Modelling* 11(1), 3–24.
- Hsieh, F., Y. Tseng, and J. Wang (2006). Joint modeling of survival and longitudinal data: likelihood approach revisited. *Biometrics* 62, 1037–1043.
- Huang, Y., X. J. Hu, and G. A. Dagne (2014). Jointly modeling time-to-event and longitudinal data: a bayesian approach. *Statistical Methods & Applications* 23(1), 95–121.
- Ibrahim, J., M.-H. Chen, and D. Sinha (2004). Bayesian methods for joint modeling of longitudinal and survival data with applications to cancer vaccine studies. *Statistica Sinica* 14(3), 863–883.
- Jiang, B., M. R. Elliott, M. D. Sammel, and N. Wang (2015, Feb). Joint modeling of cross-sectional health outcomes and longitudinal predictors via mixtures of means and variances. *Biometrics* 71(2), 487–97.
- Kneib, T. and L. Fahrmeir (2007, Mar). A mixed model approach for geoadditive hazard regression. *Scandinavian Journal of Statistics* 34(1), 207–228.
- Lang, S. and A. Brezger (2004). Bayesian p-splines. *Journal of Computational and Graphical Statistics* 13(1), 183–212.
- Lin, X., J. Raz, and S. Harlow (1997, Sep). Linear mixed models with heterogeneous within-cluster variances. *Biometrics* 53(3), 910–923.
- Lunn, D., C. Jackson, N. Best, A. Thomas, and D. Spiegelhalter (2013). *The BUGS Book: A Practical Introduction to Bayesian Analysis*. Chapman & Hall/CRC.
- Lunn, D., A. Thomas, N. Best, and D. Spiegelhalter (2000). Winbugs – a bayesian modelling framework: Concepts, structure, and extensibility. *Statistics and Computing* 10, 325–337.
- Lyles, R. H., A. Muñoz, J. Xu, J. M. G. Taylor, and J. S. Chmiel (1999). Adjusting for measurement error to assess health effects of variability in biomarkers. *Statistics In Medicine* 18, 1069–1086.
- May, M. T. and Gompels, M., V. Delpech, K. Porter, C. Orkin, S. Kegg, and C. Sabin (2014). Impact on life expectancy of hiv-1 positive individuals of cd4+ cell count and viral load response to antiretroviral

- therapy. *AIDS* 28(8), 1193–1202. London, England.
- McLain, A. C., K. J. Lum, and R. Sundaram (2012, Feb). A joint mixed effects dispersion model for menstrual cycle length and time-to-pregnancy. *Biometrics* 68(2), 648–656.
- Rizopoulos, D. (2012). *Joint Models for Longitudinal and Time-to-Event Data With Applications in R*. Chapman and Hall/CRC.
- Rizopoulos, D. and P. Ghosh (2011). A bayesian semiparametric multivariate joint model for multiple longitudinal outcomes and a time-to-event. *Stat. Med.* 30(12), 1366–1380.
- Rizopoulos, D., G. Verbeke, and E. Lesaffre (2009). Fully exponential laplace approximations for the joint modelling of survival and longitudinal data. *Journal of the Royal Statistical Society, Series B* 71, 637–654.
- Song, X. and C. Wang (2008, Jun). Semiparametric approaches for joint modeling of longitudinal and survival data with time-varying coefficients. *Biometrics* 64(2), 557–566.
- Souza-Jr, P., C. L. Szwarcwald, and E. A. Castilho (2007). Delay in introducing antiretroviral therapy in patients infected by HIV in Brazil, 2003-2006. *Clinical Science* 62(5), 579–584.
- Spiegelhalter, D., N. Best, B. Carlin, and A. Van der Linde (2002). Bayesian measures of model complexity and fit (with discussion). *Journal of the Royal Statistical Society (Series B)* 64(4), 583–639.
- Tang, N.-S., A.-M. Tang, and D.-D. Pan (2014, September). Semiparametric bayesian joint models of multivariate longitudinal and survival data. *Computational Statistics & Data Analysis* 77, 113–129.
- Tseng, Y. K., F. S. Hsieh, and J. L. Wang (2005). Joint modelling of accelerated failure time and longitudinal data. *Biometrika* 92(3), 587–603.
- Watanabe, S. (2010). Asymptotic equivalence of Bayes cross validation and widely applicable information criterion in singular learning theory. *J. Mach. Learn. Res.* 11(455), 3571–3591.
- Wulfsohn, M. and A. Tsiatis (1997). A joint model for survival and longitudinal data measured with error. *Biometrics* 53, 330–339.
- Yu, Z., L. Liu, D. M. Bravata, and L. S. Williams (2014, Nov). Joint model of recurrent events and a terminal event with time-varying coefficients. *Biometrical Journal* 56(2), 183–197.
- Zhu, H., J. Ibrahim, Y.-Y. Chi, and N. Tang (2012, September). Bayesian influence measures for joint models for longitudinal and survival data. *Biometrics* 68(3), 954–964.

Table 1: WAIC values for the 33 Bayesian joint dispersion models with flexible links.

| Longitudinal model | | Survival model | | | |
|--------------------|--------------|--------------------------------------|-------------------|--|-------------------|
| m_i | σ_i^2 | $\varrho_i(t)$ | h_0 | | |
| | | | Weibull | P-Spline | Piecewise |
| (17) | (18) | (22) + (23) | 14671 | 12573 \diamond | 14317 |
| (17) | (19) | | 14700 | 12848 | 14483 |
| (17) | (18) | (22) + (24) | 14307 | 12605 \diamond | 13365 |
| (17) | (19) | | 14452 | 12917 | 13571 |
| (17) | (20) | | 13134 \triangle | 12104 \blacktriangle | 12921 \triangle |
| (17) | (21) | | 13956 \square | 12887 \square | 13533 \square |
| (17) | (18) | (22) + (25) | 14811 | 13334 \diamond | 14463 |
| (17) | (19) | | 14923 | 13688 | 14599 |
| (17) | (20) | | 14314 | 13144 | 13968 |
| (17) | (21) | | 14627 \circ | 13553 \circ | 14355 \circ |
| (17) | (21) | (22) + $g_1 b_{1i,1} + g_2 b_{1i,2}$ | 16984 \diamond | 15779 \blacklozenge | 16383 \diamond |

Table 2: Posterior parameters estimates and 95% Credible Intervals (CI) for the selected model \blacktriangle and for the traditional joint model \blacklozenge with a baseline adjusted with P-Splines.

| Parameter | Model \blacktriangle | | Model \blacklozenge | |
|----------------------------|------------------------|----------------|-----------------------|----------------|
| | Mean | 95% CI | Mean | 95% CI |
| Longitudinal: | | | | |
| Intercept (β_{10}) | 17.26 | (16.59, 17.94) | 17.3 | (16.57, 18.02) |
| Time (β_{11}) | 1.74 | (1.46, 2.04) | 1.92 | (1.66, 2.19) |
| Gender (β_{12}) | -0.66 | (-1.42, -0.02) | -0.60 | (-1.42, 0.21) |
| Age (β_{13}) | -1.34 | (-2.51, -0.41) | -1.59 | (-2.58, -0.58) |
| PrevOI (β_{14}) | -1.62 | (-2.23, -0.99) | -1.80 | (-2.77, -0.81) |
| σ_{11}^b | 20.85 | (17.83, 24.22) | 24.58 | (21.32, 28.24) |
| σ_{22}^b | 4.71 | (3.80, 5.78) | 6.12 | (5.03, 7.36) |
| σ_{12}^b | -3.07 | (-4.46, -1.77) | -4.15 | (-5.77, -2.67) |
| σ_0 | — | — | 2.59 | (2.50, 2.67) |
| Survival: | | | | |
| Gender (β_{31}) | 0.74 | (0.49, 1.01) | 0.49 | (-0.30, 1.29) |
| Age (β_{32}) | 0.99 | (0.72, 1.27) | 0.93 | (0.04, 1.76) |
| PrevOI (β_{33}) | 1.04 | (0.80, 1.28) | 0.95 | (0.19, 1.67) |
| g_1 | — | — | -0.21 | (-0.31, -0.10) |
| g_2 | — | — | -0.54 | (-0.76, -0.33) |

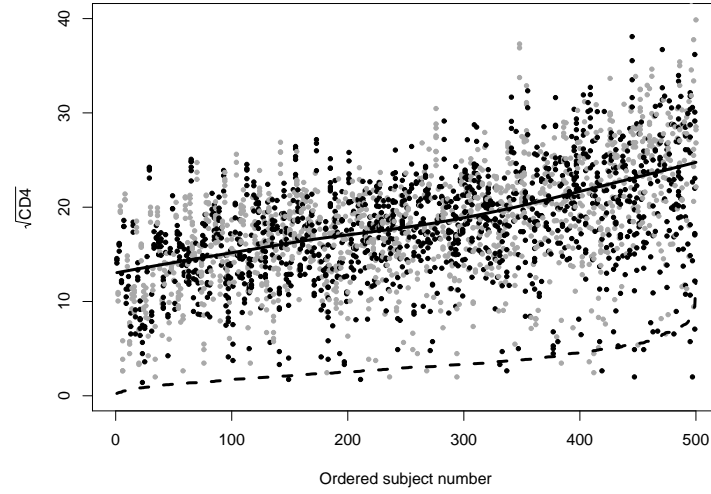


Figure 1: Plot of the $\sqrt{\text{CD4}}$ values against the subject number resulting from ordering them according to the standard deviation of their $\sqrt{\text{CD4}}$. The $\sqrt{\text{CD4}}$ values for one subject are plotted using the same color and two adjacent subjects have different colors. The solid line represents the *lowess* smooth of the $\sqrt{\text{CD4}}$ values and the dashed one is simply the connection between the 500 standard deviations.

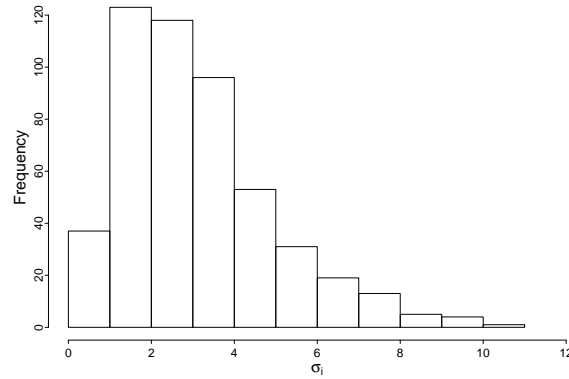


Figure 2: Histogram of the posterior means for the σ_i 's (model \blacktriangle). There's a considerable heterogeneity among these right skewed estimates.

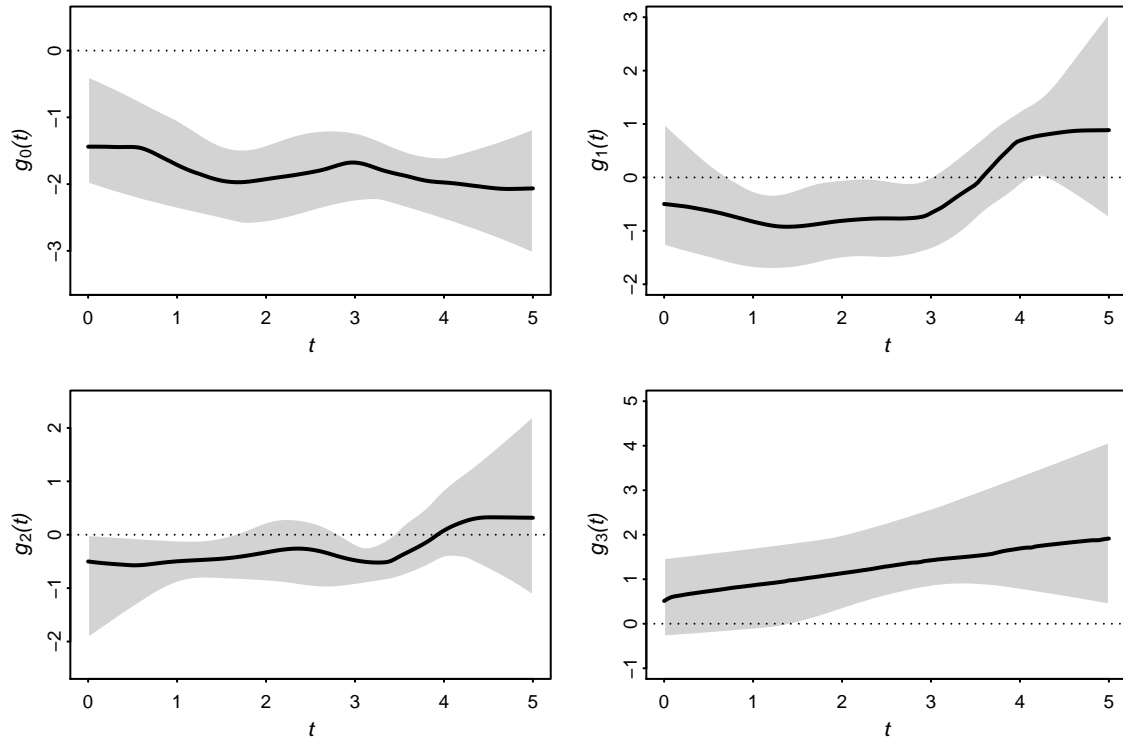


Figure 3: Posterior mean estimates, together with the corresponding 95% Credible Bands (CB), for the selected model \blacktriangle . The top left panel shows $g_0 = \log(h_0)$ and the subsequent panels have the time-varying regression coefficients as a function of time in years, t .

6 Supplementary Material

In this supplementary material we provide the detailed version for calculating the WAIC measure (section 6.1) and the BUGS code for fitting the selected joint model **▲** proposed in the paper (section 6.2).

6.1 Watanabe-Akaike information criterion (WAIC)

Following [Gelman et al. \(2014\)](#) we define WAIC as

$$\begin{aligned}
 \text{WAIC} &= -2 [\text{lppd} - p_{\text{WAIC}}] \\
 &= -2 [\log \text{ pointwise predictive density} - \text{Penalization}] \\
 &= -2 \left[\log \left\{ \prod_{i=1}^N p(\mathcal{D}_i | \mathcal{D}) \right\} - \sum_{i=1}^N \mathbb{V}_{\boldsymbol{\theta} | \mathcal{D}} [\log \{p(\mathcal{D}_i | \boldsymbol{\theta})\}] \right] \\
 &= -2 \left[\sum_{i=1}^N \log \left\{ \int p(\mathcal{D}_i | \boldsymbol{\theta}) \pi(\boldsymbol{\theta} | \mathcal{D}) d\boldsymbol{\theta} \right\} - \sum_{i=1}^N V_{s=1}^S [\log \{p(\mathcal{D}_i | \boldsymbol{\theta}^s)\}] \right] \\
 &\approx -2 \left[\sum_{i=1}^N \log \left\{ \frac{1}{S} \sum_{s=1}^S p(\mathcal{D}_i | \boldsymbol{\theta}^s) \right\} - \sum_{i=1}^N \frac{1}{S-1} \sum_{s=1}^S \left(\log \{p(\mathcal{D}_i | \boldsymbol{\theta}^s)\} - \left\{ \frac{1}{S} \sum_{s=1}^S \log p(\mathcal{D}_i | \boldsymbol{\theta}^s) \right\} \right)^2 \right] \\
 &= -2 \left[\sum_{i=1}^N \log \left\{ \overline{p(\mathcal{D}_i | \boldsymbol{\theta})} \right\} - \sum_{i=1}^N \frac{1}{S-1} \sum_{s=1}^S \left(\log \{p(\mathcal{D}_i | \boldsymbol{\theta}^s)\} - \overline{\log \{p(\mathcal{D}_i | \boldsymbol{\theta})\}} \right)^2 \right] \quad (26)
 \end{aligned}$$

where $V_{s=1}^S[a_s]$, represents the posterior sample variance and the over line stands for the posterior sample mean. Like the DIC models with smaller WAIC values should be preferred.

6.2 BUGS code

We provide here the BUGS code for fitting the selected joint model **▲**. The code can be easily generalized to other structures both on the longitudinal and survival parts.

```
#####
#### JOINT DISPERSION MODEL WITH A FLEXIBLE LINK
## Longitudinal model --> linear with subject-specific random effects (eq. 17)
## Dispersion model --> exchangeable prior for the individual variance (eq. 20)
## Hazard model --> (eq. 4)
## Baseline hazard --> PB-Splines
## Linking structure --> (eq. 24)
## linking parameters g_1(t), g_2(t) and g_3(t) --> time-dependent modeled via PB-Splines
```

```

## Penalization through a random walk of order 1.

### Nomenclature
# N - Total number of individuals
# n.exams - Total number of clinical exams per individual
# gender, age and prevoi - covariates
# Y - \sqrt{CD4}
# tauy - precision of the normal distribution
# times - individual observed measurement times
# beta1 - population parameters for the longitudinal model (\beta_1)
# b - individual random effects
# b0 and tau --> respectively, prior mean and variance for the random-effects
# status - failure indicator, \delta_i
# obs.t <- observed times, T_i
# BS.surv.time - matrix where each line corresponds to each observed time, T_i, evaluated
# for each BSpline basis function
# n.basisfunc - number of BSpline basis functions (Q)
# beta3 - \beta_3
# gamma.0, gamma.1, gamma.2 and gamma.3 --> \gamma coefficients in eq. (5)
# eta_i --> eq. 22 + eq. 24

model{

C <- 10000 # constant to use in the zeros trick

#####
### Longitudinal model
#####
for(i in 1:N){
  for(j in 1:n.exams[i]) {
    Y[i,j] ~ dnorm(muy[i,j], tauy[i])
    muy[i,j] <- beta1[1] + beta1[2]*times[i,j] + beta1[3]*gender[i] + beta1[4]*age[i] + beta1[5]*prevoi[i] +
      b[i, 1] + b[i,2]*times[i,j]

    # log(L_1) = Log-Likelihood longitudinal part for individual i at time j
    log_like_long[i,j] <- log(sqrt(tauy[i]/6.2831)) - tauy[i]*0.5*pow(Y[i,j]-muy[i,j],2)

    # log(L_1*L_2) = full log-likelihood for individual i at time j (eq. 12)
    log_like[i,j] <- log_like_long[i,j] + log_like_surv[i]
  }

  # Random effects
  b[i,1:2] ~ dnmnorm(b0[], tau[,])
}

#####
### Survival model
#####

## Calculate \varrho_i(t) at T_i (eq.22 + eq. 24)
for(i in 1:N) {
  varrho[i] <- beta3[1]*gender[i] + beta3[2]*age[i] + beta3[3]*prevoi[i] +
    g1[i]*b[i,1] + g2[i]*b[i,2] + g3[i]*sigmay[i]
}

```



```

## Calculate the value of the splines g0(.)=log[h_0], g1(.), g2(.) and g3(.)
## at T_i (obs.t[i]) --> g0[i], g1[i], g2[i] and g3[i]
for(k in 1:n.basisfunc){
  BS.g0[i, k] <- gamma.0[k] * BS.surv.time[i, k]
  BS.g1[i, k] <- gamma.1[k] * BS.surv.time[i, k]
  BS.g2[i, k] <- gamma.2[k] * BS.surv.time[i, k]
  BS.g3[i, k] <- gamma.3[k] * BS.surv.time[i, k]
}
g0[i] <- sum(BS.g0[i, 1:n.basisfunc])      ## log-baseline hazard --> log[h_0] = g_0(t)
h0[i] <- exp(g0[i])                        ## baseline hazard
g1[i] <- sum(BS.g1[i, 1:n.basisfunc])
g2[i] <- sum(BS.g2[i, 1:n.basisfunc])
g3[i] <- sum(BS.g3[i, 1:n.basisfunc])

log.hazard[i] <- (g0[i] + eta[i])*varrho[i]
}

## Calculate the integral of the hazard function, I=\int_0^{T_i}\{ \exp\{g_0(u)+\varrho_i(u) du \} }
# Using the Gauss-Kronrod quadrature rule
# phi - the 15 Gauss-Kronrod quadrature points
# phi <- c(-0.949107912342758524526189684047851, -0.741531185599394439863864773280788,
#          -0.405845151377397166906606412076961, 0, 0.405845151377397166906606412076961,
#          0.741531185599394439863864773280788, 0.949107912342758524526189684047851,
#          -0.991455371120812639206854697526329, -0.864864423359769072789712788640926,
#          -0.586087235467691130294144838258730, -0.207784955007898467600689403773245,
#          0.207784955007898467600689403773245, 0.586087235467691130294144838258730,
#          0.864864423359769072789712788640926, 0.991455371120812639206854697526329)
# w - the 15 Gauss-Kronrod quadrature weights
# w <- c(0.063092092629978553290700663189204, 0.140653259715525918745189590510238,
#        0.190350578064785409913256402421014, 0.209482141084727828012999174891714,
#        0.190350578064785409913256402421014, 0.140653259715525918745189590510238,
#        0.063092092629978553290700663189204, 0.022935322010529224963732008058970,
#        0.104790010322250183839876322541518, 0.169004726639267902826583426598550,
#        0.204432940075298892414161999234649, 0.204432940075298892414161999234649,
#        0.169004726639267902826583426598550, 0.104790010322250183839876322541518,
#        0.022935322010529224963732008058970)

# loop for the Gauss-Kronrod quadrature rule
for(i in 1:N){
  # P1=(b-a)/2 = (obs.t[i] - 0)/2 and P2=(obs.t[i] + 0)/2
  # in this case P1=P2 because the lower bound of the integral is 0=a
  P1[i] <- obs.t[i]/2
  P2[i] <- P1[i]
  # loop to calculate the value of the splines g0, g1, g2 and g3 at the 15 quadrature points, "phi"
  # u is the value (obs.t_i/2)*phi + obs.t_i/2
  # BS.u.vect is a matrix which dim=(15*500)x(n.basisfunc);
  # each line is the transformation of the quadrature points (phi) to the B-Spline Basis
  # varrho.u is the value of varrho at point u
  for (j in 1:15) {
    for(k in 1:n.basisfunc){

```

```

        BS.g0.u[i, j, k] <- gamma.0[k] * BS.u.vect[i*15-15+j, k]
        BS.g1.u[i, j, k] <- gamma.1[k] * BS.u.vect[i*15-15+j, k]
        BS.g2.u[i, j, k] <- gamma.2[k] * BS.u.vect[i*15-15+j, k]
        BS.g3.u[i, j, k] <- gamma.3[k] * BS.u.vect[i*15-15+j, k]
    }
    g0.u[i, j] <- sum(BS.g0.u[i, j, 1:n.basisfunc])
    g1.u[i, j] <- sum(BS.g1.u[i, j, 1:n.basisfunc])
    g2.u[i, j] <- sum(BS.g2.u[i, j, 1:n.basisfunc])
    g3.u[i, j] <- sum(BS.g3.u[i, j, 1:n.basisfunc])

    varrho.u[i,j] <- beta3[1]*gender[i] + beta3[2]*age[i] + beta3[3]*prevoi[i] +
        g1.u[i, j]*b[i,1] + g2.u[i, j]*b[i,2] + g3.u[i, j]*sigmay[i]

    w.exp.g0.varrho[i, j] <- w[j] * exp(g0.u[i, j] + varrho.u[i, j]) # terms to sum

}

log.survival[i] <- -P1[i] * sum(w.exp.g0.varrho[i, 1:15])
# log-Survival;  $-\int_0^{\infty} \{e^{-\{g_0(u)+\text{varrho}_i(u)\}} du\}$  } aproximates the minus integral, i.e.,
# the (-cumulative hazard), -H, summing over the 15 quadrature points

log_like_surv[i] <- log.hazard[i] + log.survival[i] # log(L2) = Log-Likelihood survival part

# To define the likelihood of the survival model we use the zeros trick
# of WinBUGS, where C is a positive constant
zeros[i] <- 0
zeta[i] <- - log_like_surv[i] + C
zeros[i] ~ dpois(zeta[i])
}

#####
### Priors

## Penalized prior for the coefficients gamma.0, gamma.1, gamma.2 and gamma.3
## of the functions g0, g1, g2 and g3
# random walk prior of order 1
gamma.0[1] ~ dnorm(0, 0.01)
gamma.1[1] ~ dnorm(0, 0.01)
gamma.2[1] ~ dnorm(0, 0.01)
gamma.3[1] ~ dnorm(0, 0.01)
for(k in 2:n.basisfunc) {
    gamma.0[k] ~ dnorm(gamma.0[k-1], tau.g0)
    gamma.1[k] ~ dnorm(gamma.1[k-1], tau.g1)
    gamma.2[k] ~ dnorm(gamma.2[k-1], tau.g2)
    gamma.3[k] ~ dnorm(gamma.3[k-1], tau.g3)
}

## prior for the individual variance
for(i in 1:N){
    log.sigmay[i] ~ dunif(-100,100)
    sigmay2[i] <- pow(exp(log.sigmay[i]), 2)
    tauy[i] <- 1/sigmay2[i]
    sigmay[i] <- sqrt(sigmay2[i]) # std. deviation of the longitudinal measures for the i individual
}

```

```

## other priors
tau.g0 ~ dgamma(0.01, 0.01)
tau.g1 ~ dgamma(0.01, 0.01)
tau.g2 ~ dgamma(0.01, 0.01)
tau.g3 ~ dgamma(0.01, 0.01)
for(k in 1:5) { beta1[k] ~ dnorm(0, 0.01) }
for(k in 1:3) { beta3[k] ~ dnorm(0, 0.01) }
tau[1:2,1:2] ~ dwish(V[,], 25)
}
# end of the model

```